

PROSPECT

Neural Stem Cells and Neuro-Oncology: Quo Vadis?

Lawrence Recht,* Taichang Jang, Todd Savarese, and N.S. Litofsky

Departments of Neurology and Surgery (Neurosurgery), University of Massachusetts Medical School, Worcester, Massachusetts 01655

Abstract Conventionally, gliomas are assumed to arise via transformation of an intraparenchymal glial cell that forms a mass that then expands centrifugally, eventually invading surrounding tissues. We propose an alternative model in which gliomas arise via initiation and promotion of cells within the brain's subependymal layer or subventricular zone, the source of a recently characterized pool of neural cells with the properties of self-renewal and multipotentiality (i.e., stem cells) that persists into adulthood. In this model, the particular histological subtype of glioma would represent the effects of temporal and spatial environmental influences rather than the particular cell of origin and the disease's centrifugal point would be the subependymal layer. The implications of such a model are discussed. *J. Cell. Biochem.* 88: 11–19, 2003. © 2002 Wiley-Liss, Inc.

Key words: neural stem cell; subependymal layer (SEL); subventricular zone (SVZ); glioma

THE CLINICAL PROBLEM

Brain tumors of glial histologies (gliomas) are the most common type of primary CNS neoplasms encountered clinically. Unfortunately, it has almost become a mantra for both clinicians and researchers to start any presentation on this topic with the fact that they remain difficult to treat despite intense research. The sad truth remains that although we have learned much about the basic biology of these tumors, little clinical progress has been made in increasing patient survival since the landmark BTSG studies over 20 years ago [Walker et al., 1978, 1980].

The discovery and characterization of a pool of cells that persists into adulthood and retains the capability to self-renew and differentiate, i.e., neural stem cells, potentially offers new ways for brain tumor researchers to approach their studies. To date, these new neurobiologic findings have been applied in two ways: (1) towards better classification of tumors, so as

hopefully to improve prognostics and identify patients who may be better suited to certain types of therapy [Noble and Dietrich, 2002]; and (2) as a potential vehicle to improve delivery of therapeutic agents into tumors [Aboody et al., 2000].

We believe that a better application of the rapidly expanding field of neural stem cell biology is to investigate the early events that occur *in situ* before macroscopic tumors develop. We propose here a model in which gliomas arise from undifferentiated stem cells whose eventual fate is influenced by local factors in a manner similar to their normal counterparts. This model provides a framework from which investigators can begin to study heretofore neglected areas of glioma, including early diagnosis and prevention, as well as providing new potential therapeutic strategies.

How Do Gliomas Arise?

Reya et al. [2001] in a recent review made the cogent observation that although much is known about the genetic mutations that fuel the cancer process at the molecular level, we frequently do not know what the effects of these lesions are in the target cell in its own environment. Phrased differently, we know much about the *molecular* but not the *cellular* biology of solid tumors, glioma included.

Consider the following assumptions that guide brain tumor researchers. It is generally

*Correspondence to: Lawrence Recht, MD, Department of Neurology, UMass Medical School, 55 Lake Av N, Worcester, MA 01655.

E-mail: Lawrence.Recht@umassmed.edu

Received 29 March 2002; Accepted 2 April 2002

DOI 10.1002/jcb.10208

© 2002 Wiley-Liss, Inc.

believed that glial tumors develop from a transformed glial, oligodendroglial or precursor cell. This cell escapes local control and begins proliferating aberrantly. In some cases, the growth is relatively slow, thus resulting in lower grades of glioma. In others, it occurs more rapidly, thus forming glioblastomas. This progression can occur stepwise, i.e., patients can first develop low-grade tumors that eventually become higher grade.

In this widely accepted model, tumors are therefore monoclonal, i.e., they develop from a single transformed cell and growth is *centrifugal*, i.e., cells spread outward as the mass enlarges and eventually invade the surrounding parenchyma. Consistent with this model is the observation that gliomas almost always arise in one intraparenchymal brain site. Furthermore, numerous instances exist of a patient having undergone a normal computed tomographic (CT) or magnetic resonance (MR) scan a few months before a large tumor is noted; this too, suggests that growth is centrifugal and arises from one site.

However, despite these examples, it is important to note that it has never been proven that gliomas arise from a resting parenchymal cell that undergoes transformation. This is because resolution of this issue requires that tumors be studied at very early time points in their development. Such study is obviously nearly impossible to accomplish using clinical brain-tumor specimens. To circumvent this, therefore, comparisons are made between established low- and high-grade gliomas on the one hand and normal brain on the other and inferences are made based on the differences. While the correlation of the differences between established tumors and normal surrounding tissue can provide some insights into early events, such information does not allow for study of what is happening at the critical moment that tumors develop.

Are there any other conceptual models that might fit the available information concerning the development of gliomas? One paradigm that has been very useful in better understanding epithelial cancers is the multistage model that was first proposed to describe the development of skin cancers after exposure to chemical carcinogens [Berenblum and Shubik, 1947, 1949] and since applied to other epithelial cancers, such as bladder, liver, and colon. This model posits that cancer develops in three stages: (1) an *initiation* stage, that presumably occurs through an irre-

versible or stable damage to cellular DNA (i.e., a genotoxic insult), (2) a *promotion* phase, which is an operational process that brings about the clonal expansion of initiated cells, and (3) a *progression* phase, which results when genetic instability leads to further mutagenic and epigenetic changes [Foulds, 1965; Trosko, 2001; Trosko and Chang, 2001].

In this model, the emergence of tumors from liver and skin after chemical carcinogenesis is explained by the initiation of pluripotent stem cells that are then suppressed by normal surrounding cells through inhibitory influences [Trosko and Chang, 2001]. Therefore, the early steps of promotion presumably result from a loss of intercellular communication that leads to the clonal multiplication and protection from apoptosis of single initiated cells.

Evaluating the Relationship of Neural Stem Cells and Glioma Using the Multistage Model

It is important to note, considering that the multistage model derived from studies of chemical carcinogenesis, that chemical exposure has been associated with brain-tumor development in epidemiologic studies [Selikoff and Hammond, 1982; Inskip et al., 1995; Carozza et al., 2000]. Furthermore, a model in which tumors emerge from initiation and promotion of a pluripotent stem cell seems an optimal framework with which someone with knowledge of the neural stem cell system could approach glioma.

To effectively link neural stem cells with glioma, however, there is a need to be able to study tumors at a very early stage. While not possible clinically, there is an experimental model of chemical neurocarcinogenesis that is ideally suited to this purpose. First described over three decades ago by Druckrey and colleagues [Druckrey et al., 1966, 1970; Ivankovic and Druckrey, 1968; Druckrey, 1971], it involves exposing rats during late gestation to a single dose of *N*-ethyl-*N*-nitrosourea (ENU). This results in the eventual development of neural tumors in virtually all offspring after a latency of greater than 4 months. Tumors are of variable types including mixed oligoastrocytomas, oligodendrogliomas, astrocytomas, and neurinomas [Zook et al., 2000]. Initiation occurs rapidly since ENU is cleared within minutes [Swann and Magee, 1971; Müller and Rajewsky, 1983] and transformation of brain cells isolated from exposed rats in the neonatal period has been reported [Laerum and Rajewsky, 1975;

Roscoe and Claisse, 1978]. The rapid clearance of ENU also allows the investigator to be fairly certain that any early changes observed are likely related to tumor development and not inflammation (which might be a problem with other models of chemical neurocarcinogenesis such as methylnitrosourea administration to adult rats or placing carcinogenic pellets in mouse brain [Zimmerman and Arnold, 1941; Seyfried, 2001]).

Several studies have assessed pathologic changes during the latency period between ENU administration and tumor development [Lantos and Cox, 1976; Schiffer et al., 1978, 1980; Lantos and Pilkington, 1979; Pilkington and Lantos, 1979; Mennel and Simon, 1985; Yoshino, 1985; Yoshino et al., 1985a,b; Ikeda et al., 1989]. As would be predicted in the multistage model where precursor lesions are observed before macroscopic tumors are noted [Farber, 1976; Solt and Farber, 1976], hyperplastic lesions with characteristics of early neoplasias could be detected several weeks before tumors are noted. These lesions were most frequently noted near the subependymal plate (at the time of these studies, this was still considered primarily a vestigial organ of unclear postnatal function and now is called the subependymal layer or subventricular zone), leading some to propose this area as a source of tumor cells [Lantos and Cox, 1976; Pilkington and Lantos, 1979].

Although detected on morphological grounds, these studies could not characterize these lesions using immunohistochemical markers. Thus, although macroscopic tumors contained cells that expressed GFAP, vimentin and Leu7 (a marker expressed on oligodendroglial cells), only one report noted the presence of the latter marker in what was deemed an early lesion [Galloway et al., 1990]. Therefore, the source of these early lesions remained uncertain.

Applying Concepts From Neural Stem Cell Biology to Neurocarcinogenesis

Farber [1976] proposed that morphology was the key to understanding early lesions and that the linkage depended on identifying markers that were present early in the course of carcinogenesis and persisted through progression. The lack of such a marker in the ENU model made linkage of these early hyperplastic lesions with macroscopic tumors somewhat tenuous. Furthermore, although the subepen-

dymal plate was a presumed source of tumor cells, its vestigial nature postnatally made it unclear what its role could be in driving the neoplastic process.

While the ENU model continues to be utilized by investigators [Blass-Kampmann et al., 1998; Adey et al., 2000; Zook et al., 2000; Kish et al., 2001], it is mostly to study the events that occur at initiation (i.e., genetic mutations) or progression (after large tumors are apparent). Little work is currently ongoing that examines the latency period, however, and to date, there are no data that suggests that a promotion stage, similar to that observed in epithelial cancers, occurs in the development of glioma [Koestner, 1990].

The recent characterization of an actively proliferating pool of cells in the SEL of adult mammals capable of responding to endogenous and exogenous influences and possessing robust migratory capacity raises several questions about what roles this area could be playing in the development of brain tumors. Our own interest in this question led us to investigate whether the SEL plays any role in glioma formation after ENU exposure. Our first experiments were based upon the assumption that if, as proposed, these early lesions represented tumors that had arisen in the subependymal layer, then they should share characteristics with this region [Jang et al., 2001a]. We therefore first assessed the expression of nestin during ENU neurocarcinogenesis. Nestin is a member of a unique class of intermediate filaments that is expressed by neural progenitors during development [Hockfield and McKay, 1985; Lendahl et al., 1990]. It is still widely expressed in brain at the time of birth, but its expression becomes restricted in normal adults to the SEL of mammals, including humans [Hockfield and McKay, 1985; Lendahl et al., 1990; Bernier et al., 2000]. Its expression can be reinduced, however, in reactive glial cells after various types of trauma [Clarke et al., 1994; Frisén et al., 1995; Duggal et al., 1997; Holmin et al., 1997; Brook et al., 1999; Kaya et al., 1999; Krum and Rosenstein, 1999; Li and Chopp, 1999], and nestin has been identified in clinical tumor specimens [Dahlstrand et al., 1992; Tohyama et al., 1992].

Since nestin expression occurs preferentially in undifferentiated, reactive and neoplastic neural cells, we reasoned that it may be expressed at all times during the development of brain tumors after ENU exposure. We noted first

(Jang et al., submitted) that nestin is detected in all tumors examined in rats that were sacrificed after 120 days of age. Nestin demarcated the tumors from surrounding parenchyma much better than GFAP and nestin+ cells were much more frequently BrdU+ than GFAP+ ones. We then extended our analysis to assess nestin expression in ENU-exposed brains from birth and were able to detect nestin+ cells in ectopic locations as early as 30 days of age. These cells were noted to occur singly or in multiple cell clusters and they tended to occur proximate to the SEL or around the corpus callosum, i.e., where early lesions are typically noted [Lantos and Cox, 1976; Schiffer et al., 1978, 1980]. We did not see such single or multiple clusters when sections were stained with vimentin or GFAP, nor did we ever see such clusters in control brains.

These nestin+ cells were distinctive enough versus the background that they could be quantified. We therefore measured the number of single and multiple cell clusters over time and measured the greatest diameter of those multiple cell clusters observed. We noted that the size of the largest lesion increased with increasing age, although the median size of the lesions was not significantly different. A bit more surprising was the observation that the ratio of single to multiple cell clusters increased with increasing age. If initiated tumor cells had spread throughout the parenchyma soon after ENU exposure where they then underwent promotion; it would be predicted that the percentage of multiple cell clusters would have increased as the rat aged. Therefore, this finding is more consistent with the possibility that lesions were either involuting or that newly initiated cells continue to migrate into the parenchyma.

We also borrowed another strategy from neural stem cell biology to culture SVZ cells obtained from neonatal rats exposed to ENU. To date, culture of brain cells involved either entire brains or cerebral hemispheres that were pooled [Roscoe and Claisse, 1976, 1978; Claisse et al., 1978; Yoshida et al., 1980; Laerum and Rajewsky, 1975; Blass-Kampmann et al., 1998]; occasional transformation was noted in these studies but the character of the cell involved in this event remained unclear. Since Reynolds & Weiss' report a decade ago [Reynolds and Weiss, 1992; Reynolds et al., 1992], it has been known that SVZ/striatal stem like cells can be isolated and maintained indefinitely in an EGF-supple-

mented chemically defined media. We therefore isolated such cells from ENU- and vehicle exposed pups at birth and serially cultured individual samples [Jang et al., 2001]. We found that normal cultures could not be maintained for more than 8 weeks in culture with serial passaging, after which growth ceased. By contrast, in 2 (22%) of the nine ENU-exposed cultures, a phenotypic change occurred after 50–60 days in culture, after which cells no longer grew as neurospheres but as a monolayer [Jang et al., 2001b]. Furthermore, they continue to grow well in the chemically defined media, and require weekly passaging. Histologically, these cells all express nestin and a smaller percentage express GFAP. Interestingly, although immortalized, these lines are not tumorigenic in either nude mice or syngeneic rat brains.

At this early point in our studies, we can therefore make the following observations/conclusions: (1) a nestin+ otherwise undifferentiated cell is present at the earliest time points in tumor formation after ENU exposure and persists throughout tumor progression; and (2) SVZ cells removed at birth occasionally undergo spontaneous immortalization after in utero exposure to ENU.

While these results remain preliminary, they have several important implications that should serve as a guide to future experiments. For example, since the finding that the earliest lesions are nestin+ suggests that the source of these cells is the SEL, an area proposed by others as the site of initiation, the question becomes when do these cells migrate from this area. Since it is known that SEL cells are capable of radial migration, providing a continual source of parenchymal cells even under normal conditions [Levison et al., 1993; Levison and Goldman, 1997; Kakita and Goldman, 1999], it is important to establish whether these initiated cells are immediately migrating or whether they migrate at a later time.

It will also be important to establish why the ratio of single cells to clusters increases with age. In regards to the possibility of regression, it is notable that studies examining the development of ENU-induced trigeminal neurinomas are consistent with the possibility that a percentage of early lesions regress [Swenberg et al., 1975]. Thus, a similar process could be occurring here. On the other hand, studies using MR imaging have noted that it is very

difficult to detect early lesions [Kish et al., 2001]. When serial scans are performed after ENU exposure, generally the scan before a new large tumor is found is normal. Thus, the possibility that these lesions become visible before regressing seems remote.

Considering our pathologic results, therefore, one potential explanation that would explain all these findings is that the observed tumor represents the descendants of a *recently* migrated cell, not the first one that migrated. This would in turn suggest that the SVZ is crucial in the effective promotion of initiated cells and that most of this process occurs there. It would also be consistent with our in vitro findings that SVZ cells can be immortalized when cultured at birth, but do not attain full malignancy, i.e., they have not yet been fully promoted. Recognizing that more work needs to be performed, these findings to date are consistent with the contention that much of the period of epigenetic promotion may be occurring in the SVZ.

CLINICAL IMPLICATIONS AND QUESTIONS ARISING

The proposed sequence of events described above represents a departure from the conventional view of brain-tumor development (Table I) and leads to some interesting possibilities. First, it leads to a different explanation for the histologic variability among gliomas. Thus, if the earliest tumor cell is an undifferentiated stem cell with, if not complete multipotentiality, at least the capacity of being susceptible to environmental and temporal influences, then histology would be more a reflection of the environment and time that initiation occurs than the cell of origin. In other words, the factors that determine whether a tumor ultimately becomes an astrocytoma or an oligoastrocytoma would

reflect more the epigenetic effects on a nestin+ undifferentiated cell rather than a different cell of origin.

This in turn suggests that instead of attempting to subclassify tumors based on their final appearance, we should be asking the following very important clinical question: *Does a glioma stem cell exist?* It has been known for decades that only a proportion of tumor cells are clonogenic or form tumors when xenografted [Park et al., 1971; Fidler and Kripke, 1977; Fidler and Hart, 1982; Nowell, 1986]. Two explanations can be offered for this finding, i.e., either all tumor cells have a general low probability of dividing or that most cancer cells have only a limited proliferative potential, i.e., there must be a stem cell.

The optimal way to distinguish tumor stem cells from other tumor cells is using separation techniques with selective markers to demonstrate that tumorigenicity, often equivalent to tumor production after xenografting, occurs with only a certain cell subset. These conditions have been best worked out for leukemia, where tumorigenicity is confined to particular pools of cells that can be detected using specific markers [Bonnet and Dick, 1997; Reya et al., 2001]. Its presence in solid tumors has been much more difficult to establish and scant attention has been paid to this issue in glioma.

From our work with the ENU model to date, we would predict that the glioma stem cell would be nestin+. This is of limited help, however, since such an intracellular marker would not be helpful in planning separation strategies. Therefore, an important area for further study in this model (and for glioma in general) will be to define markers that can be used to selectively separate these cells at early stages of tumor formation. The question of whether a glioma stem cell exists is an important issue to resolve, since its presence would

TABLE I. Explanations of Common Glioma Characteristics According to Conventional and Proposed Models of Glioma Formation

	Conventional model	Proposed model
Tumor histology	Dependent on cell of origin	Dependent on epigenetic influences on common undifferentiated precursor/stem cell
Glioma stem cell	Perhaps	Yes
Centrifugal point	Intraparenchymal	SEL
Reason for incurability	Invasiveness; tumor cell resistance to treatment	SEL continued source of promoted tumor cells

imply that treatments aimed at just reducing tumor cell number without affecting the stem cell pool would ultimately fail.

Another issue that is raised from our studies on the ENU model is the sensitivity of our detection methods. Despite the sensitivity of MR scanning, early lesions that can easily be detected pathologically are not seen using conventional MR sequences such as T2 and diffusion weighted imaging. The reason for this is unclear but could be a reflection of the fact that the pulse sequences generally used are intended to pick up changes in water content and cellular density; such an approach is clearly not optimal for detecting early lesions from their pathological appearance, which suggests that their presence is associated with little cellular distortion and edema. From a clinical perspective, it would also suggest that the availability of MR imaging has given us a false sense of security in thinking we can identify all possible tumor lesions and would serve to weaken the contention that the multi-stage model cannot be applicable to the clinical situation in glioma because most often only one lesion is noted. The findings to date using the ENU model would suggest that further work needs to be done in this area to improve our detection capability.

Finally, the issue of why treatment of gliomas almost always ultimately fails needs to be addressed in the context of this model. As stated above, in our current brain tumor model in which tumors develop and spread centrifugally from some intraparenchymal point, this is usually explained by some inherent property of the tumor cell such as invasiveness or heterogeneity in treatment susceptibility. The model proposed here, that tumor cells are not only initiated but also promoted in the SVZ before migrating to the intraparenchymal destination from which they proliferate, forces a reconsideration of just where the true source of tumor cells lies, i.e., are they continually being produced in the SVZ and are local therapies failing because this area is not being addressed. In this regard, it is notable that many studies have demonstrated how responsive the normal SVZ cell is to growth factors placed in the CSF [Craig et al., 1996; Kuhn et al., 1996; Zigova et al., 1998; Fallon et al., 2000; Benraiss et al., 2001; Pencea et al., 2001]. That tumor formation and patient outcome might be influenced by intraventricular therapies would not be seriously considered using our conventional model, but

with the present one, it would certainly merit attention.

FINAL THOUGHTS

Several years ago, Cairncross [1987] wrote a short piece in which he used several similarities between astrocytoma and chronic myelogenous leukemia to suggest that the former was also a stem cell disease. He noted, however, that the "... [hematopoietic system] ... contains recognizable self-renewing stem cells whereas gliogenesis is severely limited in the adult primate brain and true stem cells have not been found." With the identification and characterization of a CNS stem cell that persists into adulthood, however, the similarities become more compelling.

We believe it is no longer purely a speculative exercise to view glioma as a disease of "brain marrow." Furthermore, hypothesizing that glioma represents a transformation of a neural stem cell within the SVZ challenges some basic assumptions about glioma, including its "centrifugality." If verified, this model provides a framework with which to understand not only the early events of glioma but would also suggest strategies that could lead to earlier diagnosis, preventative therapies, and perhaps cures for this terrible disease.

REFERENCES

- Aboody KS, Brown A, Rainov NG, Bower KA, Liu S, Yang W, Small JE, Herrlinger U, Ourednik V, Black PM, Breakefield XO, Snyder EY. 2000. Neural stem cells display extensive tropism for pathology in adult brain: Evidence from intracranial gliomas *Proc Natl Acad Sci USA* 97(23):12846–12851.
- Adey WR, Byus CV, Cain CD, Higgins RJ, Jones RA, Kean CJ, Kuster N, MacMurray A, Stagg RB, Zimmerman G. 2000. Spontaneous and nitrosourea-induced primary tumors of the central nervous system in Fischer 344 exposed to frequency-modulated microwave fields. *Cancer Res* 60:1857–1863.
- Benraiss A, Chmielnicki E, Lerner K, Roh D, Goldman SA. 2001. Adenoviral brain-derived neurotrophic factor induces both neostriatal and olfactory neuronal recruitment from endogenous progenitor cells in the adult forebrain. *J Neurosci* 21(17):6718–6731.
- Berenblum I, Shubik P. 1947. A new, quantitative approach to the study of the stages of chemical carcinogenesis in the mouse's skin. *Br J Cancer* 1:379–391.
- Berenblum I, Shubik P. 1949. An experimental study of the initiating stage of carcinogenesis, and a re-examination of the somatic cell mutation of cancer. *Br J Cancer* 3:109–118.
- Bernier PJ, Vinet J, Cossette M, Parent A. 2000. Characterization of the subventricular zone of the adult human

- brain: Evidence for the involvement of Bcl-2. *Neurosci Res* 37:67–78.
- Blass-Kampmann S, Bilzer T, Rajewsky MF. 1998. gp130^{RB13-6}-positive neural progenitor cells are susceptible to the oncogenic effect of ethylnitrosourea in prenatal rat brain. *Neuropathol Appl Neurobiol* 24:9–20.
- Bonnet D, Dick JE. 1997. Human acute myeloid leukemia is organized as a hierarchy that originates from a primitive hematopoietic cells. *Nat Med* 3:730–737.
- Brook GA, Pérez-Bouza A, Noth J, et al. 1999. Astrocytes re-express nestin in deafferented target territories of the adult rat hippocampus. *Neuro Report* 10:1007–1011.
- Cairncross JG. 1987. The biology of astrocytoma: Lessons learned from chronic myelogenous leukemia–hypothesis. *J Neurooncol* 5:99–104.
- Carozza SE, Wrensch M, Miike R, Newman B, Olshan AF, Savitz DA, Yost M, Lee M. 2000. Occupation and adult gliomas. *Am J Epidemiol* 152(9):838–846.
- Claisse PJ, Lantos PL, Roscoe JP. 1978. Analysis of *N*-ethyl-*N*-nitrosourea-induced brain carcinogenesis by sequential culturing during the latent period. II. Morphology of the tumors induced by cell cultures. *J Natl Cancer Inst* 61:391–398.
- Clarke SR, Shetty AK, Bradley JL, Turner DA. 1994. Reactive astrocytes express the embryonic intermediate neurofilament nestin. *Neuro Report* 5:1885–1888.
- Craig CG, Tropepe V, Morshead CM, Reynolds BA, Weiss S, van der Kooy D. 1996. In vivo growth factor expansion of endogenous subependymal neural precursor cell populations in the adult mouse brain. *Neurosci* 16(8):2649–2658.
- Dahlstrand J, Collins VP, Lendahl U. 1992. Expression of the Class VI intermediate filament nestin in human central nervous system tumors. *Cancer Res* 52:5334–5341.
- Druckrey H. 1971. Genotypes and phenotypes of ten inbred strains of BD-rats. *Arzneim-Forsch* 21:1274–1278.
- Druckrey H, Ivankovic S, Preussmann R. 1966. Teratogenic and carcinogenic effects in the offspring after a single injection of ethylnitrosourea to pregnant rats. *Nature* 210:1378–1379.
- Druckrey H, Landschütz C, Ivankovic S. 1970. Transplacental induction of malignant tumors of the nervous system. II. Ethylnitrosourea in 10 genetically defined strains of rats. *Z Krebsforsch* 73:371–386.
- Duggal N, Schmidt-Kastner R, Hakim AM. 1997. Nestin expression in reactive astrocytes following focal cerebral ischemia in rats. *Brain Res* 768:1–9.
- Fallon J, Reid S, Kinyamu R, Opole I, Opole R, Baratta J, Korc M, Endo TL, Duong A, Nguyen G, et al. 2000. In vivo induction of massive proliferation, directed migration, and differentiation of neural cells in the adult mammalian brain. *Proc Natl Acad Sci USA* 97(26):14686–14691.
- Solt D, Farber E. 1976. New principle for the analysis of chemical carcinogenesis. *Nature* 263:701–703.
- Fidler IJ, Hart IR. 1982. Biological diversity in metastatic neoplasms: Origins and implications. *Science* 217:998–1003.
- Fidler IJ, Kripke MI. 1977. Metastasis results from pre-existing variant cells within a malignant tumor. *Science* 197:893–895.
- Foulds L. 1965. Multiple etiologic factors in neoplastic development. *Cancer Res* 25:1339–1347.
- Frisén J, Johansson CB, Török C, Risling M, Lendahl U. 1995. Rapid, widespread, and longlasting induction of nestin contributes to the generation of glial scar tissue after CNS injury. *J Cell Biol* 131(2):453–464.
- Galloway PG, Likavec MJ, Perry G. 1990. Immunohistochemical recognition of ethylnitrosourea induced rat brain microtumors by anti-Leu 7 monoclonal antibody. *Cancer Lett* 49:243–248.
- Hockfield S, McKay RDG. 1985. Identification of major cell classes in the developing mammalian nervous system. *J Neuroscience* 5(12):3310–3328.
- Holmin S, Almqvist P, Lendahl U, Mathiesen T. 1997. Adult nestin-expressing subependymal cells differentiate to astrocytes in response to brain injury. *Eur J Neurosci* 9:65–75.
- Ikeda T, Mashimoto H, Iwasaki K, Shimokawa I, Matsuo T. 1989. A sequential ultrastructural and histoautoradiographic study of early neoplastic lesions in ethylnitrosourea-induced rat glioma. *Acta Pathol Jpn* 39:487–495.
- Inskip PD, Linet MS, Heineman EF. 1995. Etiology of brain tumors in adults. *Epidemiol Rev* 17(2):382–414.
- Ivankovic S, Druckrey H. 1968. Transplacentare Erzeugung maligner tumoren des Nervensystems. I. Äthyl-nitroso-harnstoff (ÄNH) in BD IX-ratten. *Z Krebsforsch* 71:320–360.
- Jang T, Litofsky NS, Ross A, Recht L. 2001a. Analysis of subventricular zone (SVZ) kinetics during ethylnitrosourea (ENU)-induced neuro-carcinogenesis. *J Neurooncol* 3:276.
- Jang T, Low HP, Salmonsén R, Savarese T, Hsieh C, Liu Q, Ross AH, Litofsky NS, Recht L. 2001b. In vitro transformation of SVZ-derived EGF-responsive precursor cells after isolation from neonates transplacentally exposed to ENU. *J Neurooncol* 3:276–277.
- Kakita A, Goldman JE. 1999. Patterns and dynamics of SVZ cell migration in the postnatal forebrain: Monitoring living progenitors in slice preparations. *Neuron* 23:461–472.
- Kaya SS, Mahmood A, Li Y, Yavuz E, Chopp M. 1999. Expression of nestin after traumatic brain injury in rat brain. *Brain Res* 840:153–157.
- Kish PE, Blaivas M, Strawderman M, Muraszko KM, Ross DA, Ross BD, McMahon G. 2001. Magnetic resonance imaging of ethyl-nitrosourea-induced rat gliomas: A model for experimental therapeutics of low-grade glioma. *J Neurooncol* 53(3):243–257.
- Koestner A. 1990. Characterization of *N*-nitrosourea-induced tumors of the nervous system; Their prospective value for studies of neurocarcinogenesis and brain tumor therapy. *Toxicol Pathol* 18(1 (Part 2)):186–192.
- Krum JM, Rosenstein JM. 1999. Transient coexpression of nestin, GFAP, and vascular endothelial growth factor in mature reactive astroglia following neural grafting or brain wounds. *Exp Neurol* 160:348–360.
- Kuhn HG, Dickinson-Anson H, Gage FH. 1996. Neurogenesis in the dentate gyrus of the adult rat: Age-related decrease of neuronal progenitor proliferation. *J Neurosci* 16(6):2027–2033.
- Laerum OD, Rajewsky MF. 1975. Neoplastic transformation of fetal rat brain cells in culture after exposure to ethylnitrosourea in vivo. *J Natl Cancer Inst* 55:1177–1187.

- Lantos PL, Cox DJ. 1976. The origin of experimental brain tumours: A sequential study. *Experientia* 32:1467–1468.
- Lantos PL, Pilkington GJ. 1979. The development of experimental brain tumors. A sequential light and electron microscope study of the subependymal plate. I. Early lesions (abnormal cell clusters). *Acta Neuropathol* 45:167–175.
- Lendahl U, Zimmerman LB, McKay RDG. 1990. CNS stem cells express a new class of intermediate filament protein. *Cell* 60:585–595.
- Levison SW, Goldman JE. 1997. Multipotential and lineage restricted precursors coexist in the mammalian perinatal subventricular zone. *J Neurosci Res* 48:83–94.
- Levison SW, Chuang C, Abramson BJ, Goldman JE. 1993. The migrational patterns and developmental fates of glial precursors in the rat subventricular zone are temporally regulated. *Development* 119:611–622.
- Li Y, Chopp M. 1999. Temporal profile of nestin expression after focal cerebral ischemia in adult rat. *Brain Res* 838:1–10.
- Mennel HD, Simon H. 1985. Morphology of early stages of ENU-induced brain tumors in rats. *Exp Pathol* 28:207–214.
- Müller R, Rajewsky MF. 1983. Elimination of O₆-ethylguanine from the DNA of brain, liver, and other rat tissues exposed to ethylnitrosourea at different stages of prenatal development. *Cancer Res* 43:2897–2904.
- Noble M, Dietrich J. 2002. Intersections between neurobiology and oncology: Tumor origin, treatment, and repair of treatment-associated damage. *Trends Neurosci* 25(2):103–107.
- Nowell PC. 1986. Mechanisms of tumor progression. *Cancer Res* 46:2203–2207.
- Park CH, Bergsagel DE, McCulloch E. 1971. Mouse myeloma tumor stem cells: A primary cell culture assay. *J Natl Cancer Inst* 46:411–422.
- Pencea V, Bingaman KD, Wiegand SJ, Luskin MB. 2001. Infusion of brain-derived neurotrophic factor into the lateral ventricle of the adult rat leads to new neurons in the parenchyma of the striatum, septum, thalamus, and hypothalamus. *J Neurosci* 21(17):6706–6717.
- Pilkington GJ, Lantos PL. 1979. The development of experimental brain tumors. A sequential light and electron microscope study of the subependymal plate. II. Microtumors. *Acta Neuropathol (Berl)* 45:177–185.
- Reya T, Morrison SJ, Clarke MF, Weissman IL. 2001. Stem cells, cancer, and cancer stem cells. *Nature* 414:105–111.
- Reynolds BA, Weiss S. 1992. Generation of neurons and astrocytes from isolated cells of the adult mammalian central nervous system. *Science* 255:1707–1710.
- Reynolds BA, Tetzlaff W, Weiss S. 1992. A multipotent EGF-responsive striatal embryonic progenitor cell produces neurons and astrocytes. *J Neurosci* 12(11):4565–4574.
- Roscoe JP, Claisse PJ. 1976. A sequential in vivo–in vitro study of carcinogenesis induced in the rat brain by ethylnitrosourea. *Nature* 262:314–316.
- Roscoe JP, Claisse PJ. 1978. Analysis of *N*-ethyl-*N*-nitrosourea-induced brain carcinogenesis by sequential culturing during the latent period. I. Morphology and tumorigenicity of the cultured cells and their growth in agar. *J Natl Cancer Inst* 61:381–390.
- Schiffer D, Giordana MT, Pezzotta S, Lechner C, Paoletti P. 1978. Cerebral tumors induced by transplacental ENU: Study of the different tumoral stages, particularly of early proliferations. *Acta Neuropathol* 41:27–31.
- Schiffer D, Giordana MT, Mauro A, Racagni G, Bruno F, Pezzotta S, Paoletti P. 1980. Experimental brain tumors by transplacental ENU. Multifactorial study of the latency period. *Acta Neuropathol* 49:117–122.
- Selikoff IJ, Hammond EC. 1982. Brain tumors in the chemical industry. *Ann N Y Acad Sci* 381:1–364.
- Seyfried TN. 2001. Perspectives on brain tumor formation involving macrophages, glia, and neural stem cells. *Perspect Biol Med* 44(2):263–282.
- Farber E. 1976. Putative precursor lesions: Summary and some analytical considerations. *Cancer Res* 36:2703–2705.
- Swann PF, Magee PN. 1971. Nitrosamine-induced carcinogenesis. The alkylation of N-7 of guanine of nucleic acids of the rat by diethylnitrosamine, *N*-ethyl-*N*-nitrosourea and ethyl mthanesulphonate. *Biochem J* 125:841–847.
- Swenberg JA, Clendenon N, Denlinger R, Gordon WA. 1975. Sequential development of ethylnitrosourea-induced neurinomas: Morphology, biochemistry, and transplantability. *J Natl Cancer Inst* 55(1):147–152.
- Tohyama T, Lee VMY, Rorke LB, Marvin M, McKay RDG, Trojanowski JQ. Nestin expression in embryonic human neuroepithelium and in human neuroepithelial tumor cells. *Lab Invest* 66(3):303–313.
- Trosko JE. 2001. Commentary: Is the concept of “tumor promotion” a useful paradigm? *Mol Carcinog* 30:131–137.
- Trosko JE, Chang C-C. 2001. Mechanism of up-regulated gap junctional intercellular communication during chemoprevention and chemotherapy of cancer. *Mutat Res* 480–481:219–229.
- Walker MD, Alexander E, Hunt WE, MacCarty CS, Mahaley MS, Mealey J, et al. 1978. Evaluation of BCNU and/or radiotherapy in the treatment of anaplastic gliomas. A cooperative clinical trial. *J Neurosurg* 49:333–343.
- Walker MD, Green SB, Byar DP, Alexander E, Batzdorf U, Brooks WH, et al. 1980. Randomized comparisons of radiotherapy and nitrosoureas for the treatment of malignant glioma after surgery. *New Engl J Med* 303(23):1323–1329.
- Yoshida J, Cravioto H, Ransohoff J. 1980. In vitro transformation of fetal brain cells from CDF rats exposed in utero to *N*-ethyl-*N*-nitrosourea: Morphologic and immunologic studies. *J Natl Cancer Inst* 84:1231–1239.
- Yoshino T. 1985. Morphological maturation of tumor cells induced by ethylnitrosourea (ENU) in rat brains. II. On the tumors by administration of ENU in the mid-gestational stage. *Acta Pathol Jpn* 35(6):1397–1408.
- Yoshino T, Motoi M, Ogawa K. 1985a. Immunohistochemical studies on cellular character of microtumors induced by ethylnitrosourea in the rat brain utilizing anti-Leu 7 and anti-glial fibrillary acidic protein antibodies. *Acta Neuropathol* 66:167–169.
- Yoshino T, Motoi M, Ogawa K. 1985b. Morphological maturation of tumor cells induced by ethylnitrosourea (ENU) in rat brains. I. On the tumors by administration

- of ENU in the late gestational stage. *Acta Pathol Jpn* 35(6):1385–1396.
- Zigova T, Pencea V, Wiegand SJ, Luskin MB. 1998. Intraventricular administration of BDNF increases the number of newly generated neurons in the adult olfactory bulb. *Mol Cell Neurosci* 11:234–245.
- Zimmerman HM, Arnold H. 1941. Experimental brain tumors. I. Tumors produced with methylcholanthrene. *Cancer Res* 1:919–938.
- Zook BC, Simmens SJ, Jones RV. 2000. Evaluation of ENU-induced gliomas in rats: Nomenclature, immunohistochemistry, and malignancy. *Toxicol Pathol* 28(1):193–201.